

### Hydrodynamic study of gold nanoparticles carrying protein and PEG-like filler

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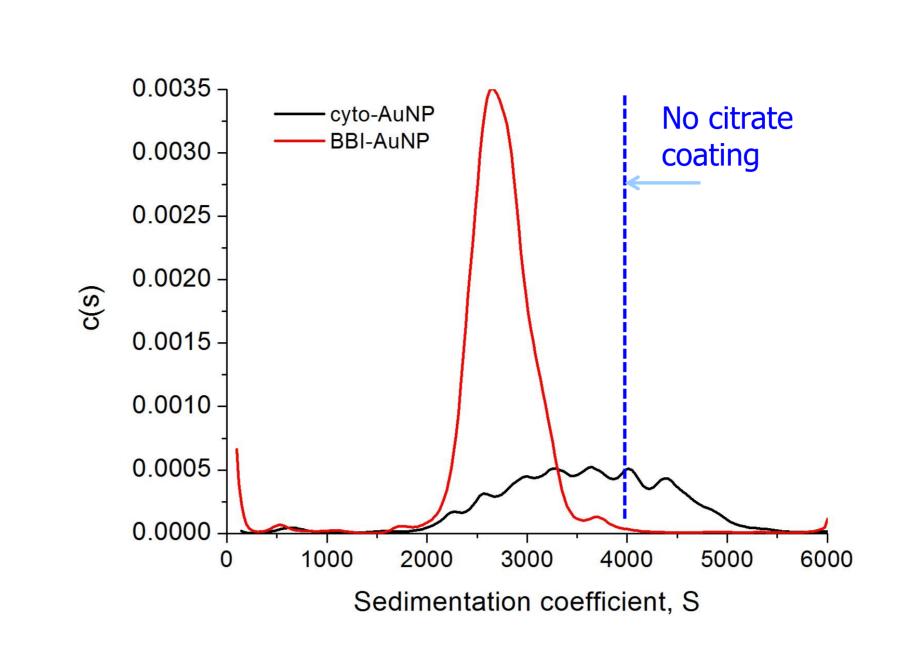
#### Introduction

Gold nanoparticles (Au-NPs) offer great potential in variety of biomedical applications such as drug delivery, cancer treatment and many others (1). AuNPs represent an attractive carriers for functional biological matrix since they are biocompatible (2) and could be manufactured in well-defined sizes. Significant progress has been made in characterisation of ligand shell for coated nanoparticles (3), however, the characterisation of Au-NP interacting with complex biological moieties still remains a challenge. In particular, the correctly oriented biological coating is the matter of a great interest. It has been shown that outer membrane  $\beta$ -barrel proteins from *E.coli* can be engineered to form self-assembled monolayer on the gold planar surfaces (4,5). In this instance the direct coupling with gold is achieved by utilizing the gold-thiol chemistry in form of addition of a single cysteine residue into one of periplasmic loops of protein  $\beta$ -barrel along with thiolAlkylPEG molecules mimicking the membrane bilayer (6)

The aim of present study was to extend the previous work on protein selfassembling onto gold planar surfaces for the case of gold nanoparticles and to characterize outer membrane protein-AuNP conjugates using classical hydrodynamic methods.

### Hydrodynamic characterization of commercial gold nanoparticles

Two types of commercial nanoparticles were characterised: cyto-AuNP (Cytodiagnostics, Canada) and BBI-AuNP (BBI Solutions, UK). Both samples demonstrate quite broad 1D distribution. Further analysis using 2D distribution shows that BBI are spherical and slightly variable only in terms of citration while cyto-AuNP had more complicated 2D distribution profiles. All consecutive experiments were carried out using **BBI** AuNP



All Au-NP were

populated with

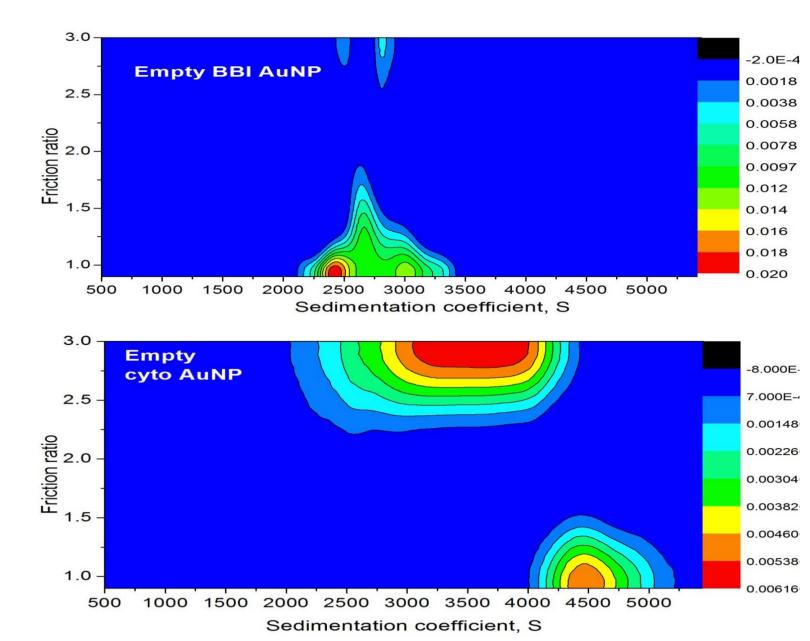
protein/protein

virtually no free

NPs were left

and filler,

0.004



— empty AuNP

+Filler

GGzOmpA-AuNP

GGzOmpA-AuNP



AuNP=Sphere with the diameter of 20 nm Volume:  $V = V_{Au} + V_{citrate\ shell} = \frac{4}{3} \frac{\pi N_A R_h^3}{\bar{v}_{particle}}$ 

Radius=Hydrodynamic Radius:  $R_h = R_h^{Au} + R_h^{citrate\ shell}$ Mass:  $M_{AuNP} = M_{AuNP} + M_{citrate\ shell} = \frac{5.11}{D(1-\bar{v}_{particle}\rho_{solvent})}$ 

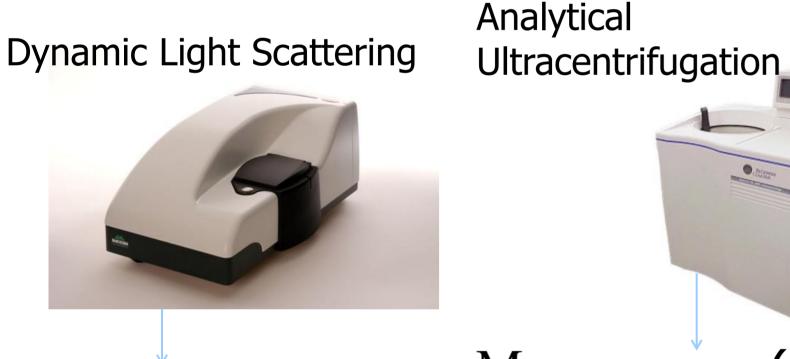
Translation Diffusion Coefficient:  $D = \frac{RT}{N_A f}$ Friction coefficient:  $f = 6\pi \eta R_h$ 

Friction ratio for a sphere  $f/f_0=1$ 

Sedimentation coefficient:  $s = \frac{M(1 - \bar{v}_{particle}\rho_{solvent})}{N_A f} = \frac{M(1 - \bar{v}_{particle}\rho_{solvent})}{6N_A \pi \eta R_h}$ 

Partial Specific volume:  $\bar{v}_{particle} = \frac{M_{Au}\bar{v}_{Au} + M_{citrate}\bar{v}_{citrate}}{M_{Au} + M_{citrate}} = \frac{(2R_h)^2}{(2R_h)^2\rho_{solvent} + 18\eta_{solvent}s}$  (8)

s, c(s), c(s,  $f/f_0$ )



 $D_t$ ,  $R_h$  $ar{v}_{particle}$ 

0.050

0.058

0.065

0.073

0.083

0.093

0

0.4

8.0

1.2

1.6

2.0

0

48,692

40,139

35,790

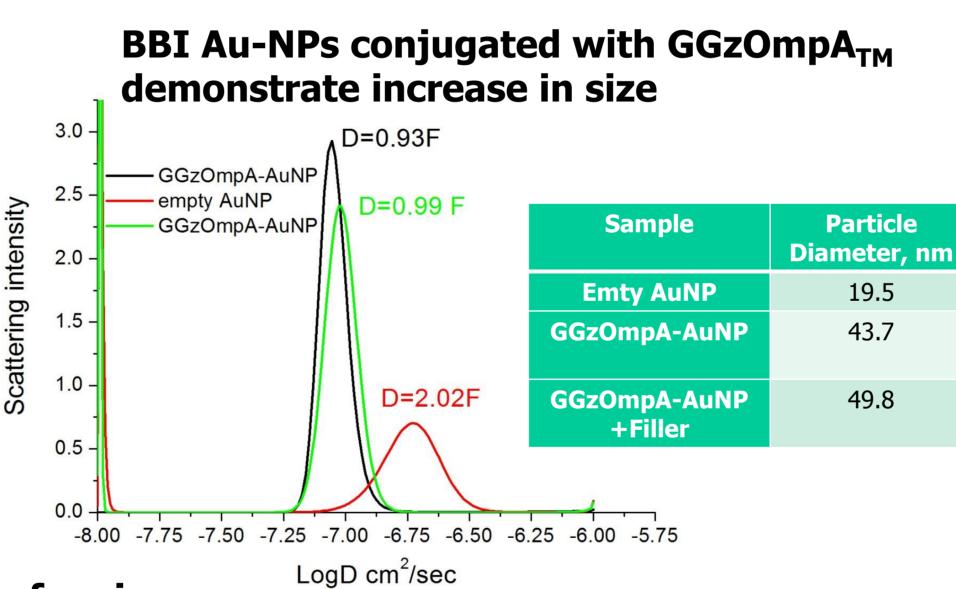
31,806

28,168

24,861

nanoparticle (7) **Z** domain protein A

**Protein assembly on the surface of gold** 



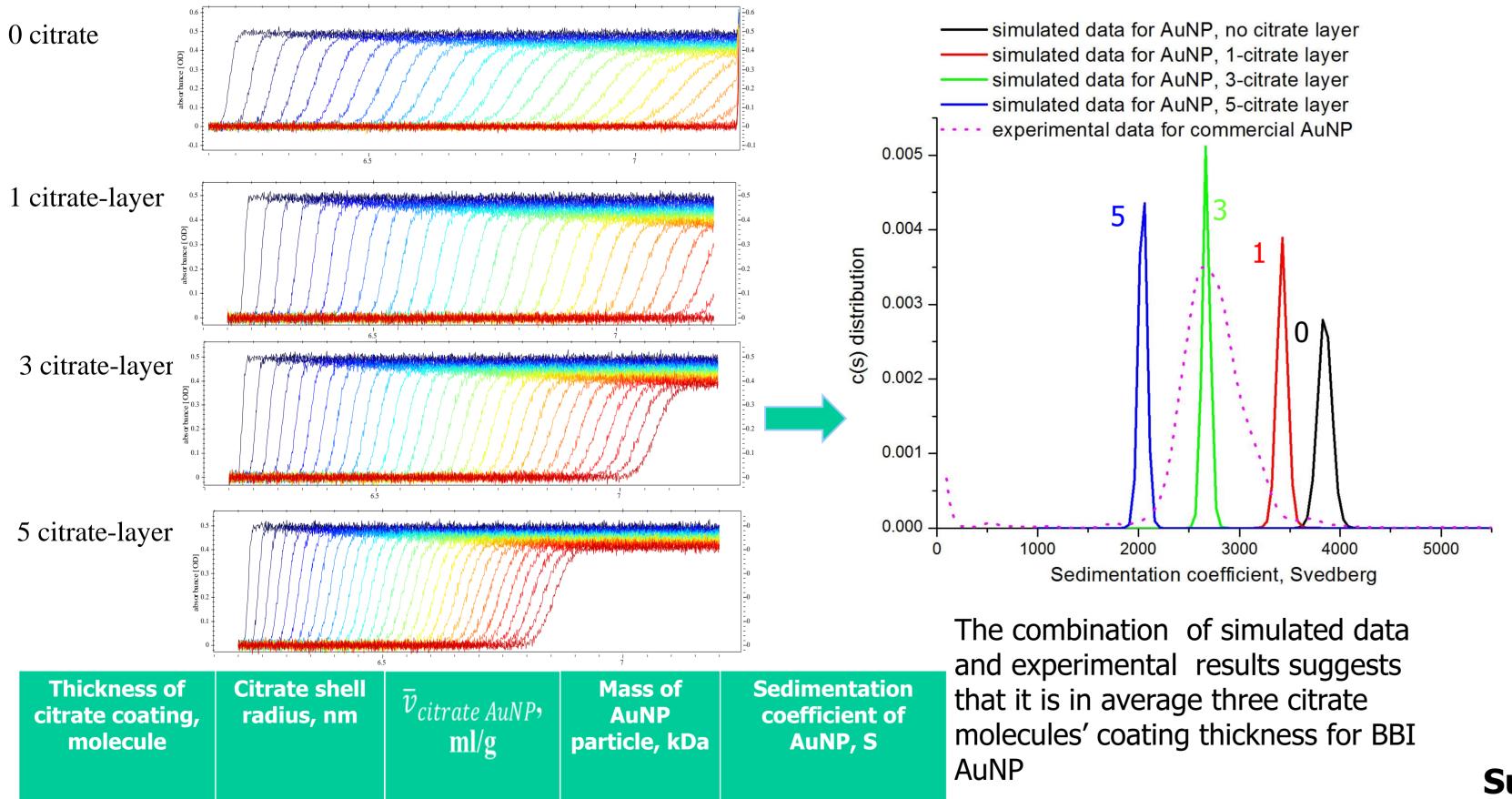
Sedimentation velocity size-distribution of BBI Au-NP coated with **GGzOmpA<sub>TM</sub>+/-filler** demonstrated changes in size and density of the particles. Filler presence resulted in some decrease in the amount of assymetrical/aggregated species

> S 0.002 2000 2500 3500 1500 3000 Sedimentation coefficient, S 2.5 -2.0 -1.5 -1500 **GGzOmpA-AuNP GGzOmpA-AuNP+Filler** 2.5 -2.0 -1.5 -3000

> > Sedimentation coefficient, S

## Simulated velocity boundaries clearly demonstrate effect of various

# thickness citrate coating to AuNP sedimentation behaviour



4007

3424

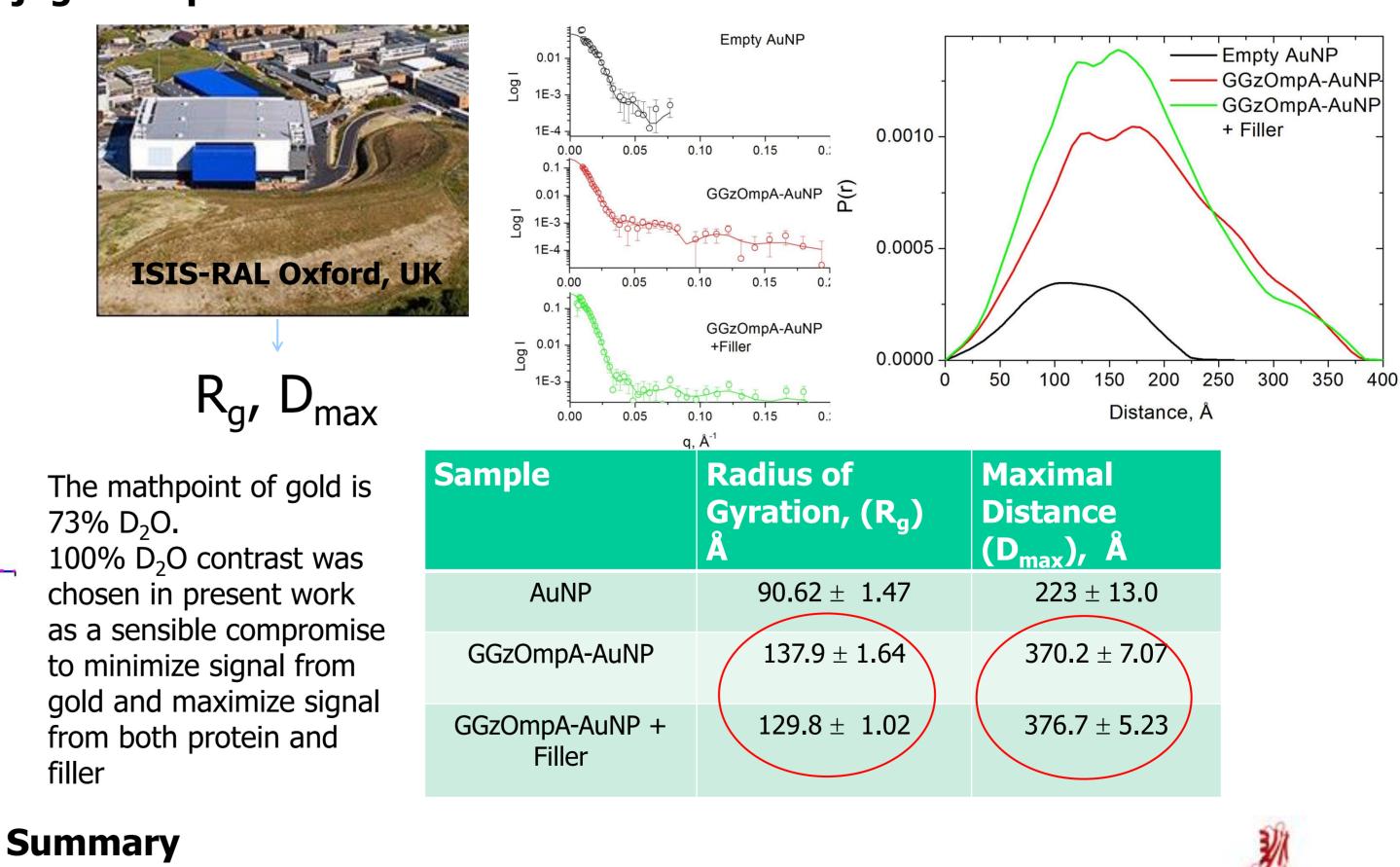
3030

2669

2334

2941

Small-Angle Neutron Scattering data suggest the formation of less asymmetrical conjugate in presence of the filler



GGzOmpA<sub>TM</sub> is effectively binding to AuNPs. The incorporation of filler molecules into the GGzOmpA<sub>TM</sub> layer results in a more ordered protein-filler array

**AuNP** 

GGzOmpA-AuNP conjugate

GGzOmpA-AuNP conjugate+PEG-like filler

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References: 1. Cabuzu, D., Cirja, A., Puiu, R, Grumezescu, A. M., Curr Top Med Chem 15, 1605 (2015). 2. Orlando, A.; Colombo, M.; Prosperi, D.; Corsi, F.; Panariti, A.; Rivolta, I.; Masserini, M.; Cazzaniga, E. J. Nanoparticle Res., 18, 58. (2016) 3. Ong, Q., Luo, Z, Stellacci, F., Accounts Of Chemical Research, 50, 1911 (2017). 4. Brun, A. P. L.; Holt, S. A.; Shah, D. S.; Majkrzak, C. F.; Lakey, J. H. Eur. Biophys. J., 37, 639–645 (2008). 5. Le Brun, A. P.; Holt, S. A.; Shah, D. S. H.; Majkrzak, C. F.; Lakey, J. H. *Biomaterials*, 32, 3303–3311 (2011). 6. Cisneros, D. A.; Muller, D. J.; Daud, S. M.; Lakey, J. H. *Angew. Chem. Int.* Ed., 45, 3252–3256 (2006) 7. Shah, D.S., Thomas M.B. Phillips, S., Cisneros D.A. Le Brun A.P. Holt S.A. Lakey J.H. Biochem. Soc. Trans. 35, 522 (2007). 8 Mächtle, W.; Borger, L. Analytical Ultracentrifugation of Polymers and Nanoparticles; Springer, (2006)